

EJP 50632

Facilitation of 8-OHDPAT-induced forepaw treading of rats by the 5-HT₂ agonist DOI

Jørn Arnt * and John Hyttel

Department of Pharmacology, H. Lundbeck A/S, Copenhagen, Denmark

Received 20 September 1988, accepted 8 November 1988

The potency of the serotonin_{1A} (5-HT_{1A}) agonist, 8-hydroxy-2-(di-n-propylamino)tetralin (8-OHDPAT), to induce forepaw treading was increased 20-fold after co-treatment with the 5-HT₂ agonist, 1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane (DOI). DOI induced head twitches which were inhibited by 8-OHDPAT. The putative 5-HT_{1B} agonist, 1-(3-trifluoromethylphenyl)piperazine (TFMPP), had a weak effect on the responses to DOI or 8-OHDPAT. The forepaw treading induced by 8-OHDPAT plus DOI was inhibited by high doses of (–)-alprenolol, ketanserin or ritanserin, but was not influenced by the β -adrenoceptor antagonist, ICI 118,551, or the 5-HT₃ antagonist, ICS 205-930. A non-effective dose of (–)-alprenolol increased the inhibitory effect of ketanserin and ritanserin. These results indicate a complex and different interaction between 5-HT_{1A} and 5-HT₂ receptors in the expression of two behavioural responses mediated by 5-HT.

5-HT_{1A} receptor agonists; 5-HT_{1B} receptor agonists; 5-HT₂ receptor agonists; Head twitches; Forepaw treading

1. Introduction

It is widely accepted that serotonin (5-HT) receptors can be divided into a number of types of which the 5-HT_{1A} and 5-HT₂ subclasses are most thoroughly characterized (Glennon, 1987). Selective agonists, e.g. 1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane (DOI; Glennon, 1987), and antagonists have been developed for 5-HT₂ receptors. In contrast, only agonists with 5-HT_{1A} selectivity are available, e.g. 8-hydroxy-2-(di-n-propylamino)tetralin (8-OHDPAT; Hjorth et al., 1982).

Various behavioural effects are mediated by 5-HT receptors in vivo, including the so-called '5-HT syndrome' (forepaw treading, head weaving, flat posture, tremor) and head twitches (Green

and Grahame-Smith, 1976; Sloviter et al., 1978; Lucki et al., 1984; Tricklebank, 1985; Glennon, 1987). Experimental evidence indicates that head twitches are mediated by 5-HT₂-receptor stimulation (Niemegeers et al., 1983; Tricklebank, 1985). It has been suggested that parts of the 5-HT syndrome are mediated by 5-HT_{1A} receptors (Tricklebank et al., 1985a). The possibility of an interaction between 5-HT_{1A} and 5-HT₂ receptors in the expression of these behaviours was suggested by Goodwin and Green (1985). However, this has not been investigated with selective agonists and was the subject of the present study.

2. Materials and methods

2.1. Animals

Male Wistar (Mol:Wist) SPF rats weighing 170–220 g were used. The rats were housed conventionally in Macrolon type III cages in groups

* To whom all correspondence should be addressed: Department of Pharmacology, H. Lundbeck A/S, Ottiliavej 7-9, DK-2500 Copenhagen-Valby, Denmark.

of 4-5 in animal rooms at $21 \pm 1^\circ\text{C}$ with a relative humidity of $55 \pm 5\%$, air exchange (16 times per hour) and a day/night cycle (light on 6 a.m.-6 p.m.). They had free access to commercial food pellets and tap water.

2.2. Behavioural observations

The rats were injected s.c. with test compounds or saline and were placed individually in Perspex observation cages (12×25 cm). Forepaw treading behaviour was scored as absent (score 0), weak or periodic (score 1) or continuous (score 2). Each rat was scored 15, 30, 45 and 60 min after injection. The scores were accumulated for each rat. Furthermore, the head twitches were counted between 30 and 40 min after injection. ED_{50} values were calculated by log-probit analysis after the effect of each dose had been expressed as per cent of the maximum possible score above the baseline scores obtained in the control groups. The maximum score for forepaw treading was 8, and 11 episodes were considered as a maximum effect for head twitches. Comparisons between groups were made by means of the Kruskal-Wallis analysis of variance followed by individual comparisons based on the Mann-Whitney U-test.

2.3. Receptor binding

[^3H]Ketanserin binding to cortical 5-HT₂ receptors was determined as follows: the tissue was homogenized (Ultra Turrax, 10 s) in 5 ml of ice-cold 50 mM Tris-buffer pH 7.7 (at 25°C). The centrifuge tubes used in this step were rinsed in ethanol by sonication for 10 min. The homogenate plus a 5 ml rinse of the homogenizer was centrifuged twice at $20\,000 \times g$ for 10 min at 4°C , with rehomogenization of the pellet in $5 + 5$ ml ice cold buffer. The final pellet was homogenized in 450 volumes (w/v) ice-cold buffer.

Triplicate incubation tubes kept on ice received 100 μl of drug solution in water (or water for total binding) and 1800 μl of tissue suspension (final tissue content corresponds to 4 mg original tissue). The binding experiment was initiated by the addition of 100 μl of [^3H]ketanserin and placing the tubes in a 37°C water bath. After incubation for

30 min the samples were filtered under vacuum (0-50 mBar) through Whatman GF/F filters (25 mm). The tubes were rinsed with 5 ml ice-cold buffer which was then poured on the filters. Thereafter, the filters were washed with 2×5 ml of buffer. The filters were placed in counting vials and PicofluorTM15, 4 ml, was added. After shaking for 1 h and 2-h storage in the dark the radioactivity was determined by liquid scintillation counting. The non-specific binding was determined in the presence of 1 μM of mianserin.

Inhibition of the binding of [^3H]8-OHDPAT or [^3H]5-HT to rat brain membranes was determined by methods analogous to the [^3H]ketanserin assay. Briefly, the rat brains were homogenized in 10 ml of 50 mM Tris buffer pH 8.0 (at 25°C) containing 120 mM NaCl, 4 mM CaCl_2 and 4 mM MgCl_2 . The homogenate was centrifuged ($20\,000 \times g$, 10 min, 4°C). The pellet was homogenized in 10 ml of the same buffer, incubated for 10 min at 37°C and centrifuged as above. The final pellet was homogenized in 100 volumes (w/v) of the buffer containing 10 μM of pargyline. Aliquots (10 mg tissue) were incubated for 15 min at 37°C with 1 nM of [^3H]8-OH-DPAT or 2 nM [^3H]5-HT (for 5-HT_{1A} and 5-HT_{1B} binding, respectively) alone or in the presence of test compound in a total volume of 1200 μl . After incubation, the samples were treated like the [^3H]ketanserin samples. Non-specific binding was obtained in the presence of 10 μM 5-HT (5-HT_{1A}) or 0.1 mM TFMPP (5-HT_{1B}).

2.4. Drugs

The following drugs were used: 8-hydroxy-2-(di-n-propylamine)tetralin hydrobromide (8-OHDPAT, molecular weight (mw) 328; synthesized by Dr. J. Perregaard in the Department of Medicinal Chemistry, H. Lundbeck A/S); 1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane hydrochloride (DOI, mw 358; Research Biochemicals Inc., USA); 1-(3-trifluoromethylphenyl)piperazine dihydrochloride (TFMPP, mw 303; obtained from Aldrich Biochemicals as free base and converted to the dihydrochloride at H. Lundbeck A/S). (–)-Alprenolol tartrate (mw 399, Hässle, Sweden); ICI 118.551 (erythro-DL-1-(7-methylindan-4-

xyloxy)-3-isopropylamino-butan-2-ol hydrochloride, mw 311, ICI, UK); ICS 205-930 (3 α -tropanyl)-1H-indole-3-carboxylic acid ester, mw 284, Sandoz, Switzerland); ketanserin, mw 395, and ritanserin, mw 478 (both Janssen, Belgium). Serotonin creatinine sulphate (Sigma, USA); [G-³H]5-hydroxytryptamine creatinine sulphate (spec. act. 18 Ci/mmol, Amersham International plc.); [ethylene-³H]ketanserin hydrochloride (spec. act. 77 Ci/mmol, New England Nuclear, FRG); [2,3(n)-³H]2-(N₁,N₁-dipropylamino)-8-hydroxy-1,2,3,4-tetrahydronaphthalene [³H]8-OHDPAT, spec. act. 200 Ci/mmol, Amersham International plc).

ICS 205-930 was dissolved in a minimum amount of 0.1 N hydrochloric acid; ketanserin and ritanserin were dissolved in a minimum amount of tartaric acid. The solutions were diluted with saline. All other compounds were dissolved in saline. The injection volumes were 5 mg/kg body weight.

3. Results

3.1. Receptor binding

The results are shown in table 1. 8-OHDPAT and (–)-alprenolol had a selective affinity for [³H]OHDPAT binding sites whereas DOI, ketanserin and ritanserin had a selective affinity for [³H]ketanserin binding sites. These compounds had weak affinities for the putative 5-HT_{1B} receptor labelled by [³H]5-HT. Finally, TFMPP had a preferential affinity for [³H]5-HT binding sites but only a marginally weaker affinity for [³H]OHDPAT and [³H]ketanserin binding sites.

3.2. Effects of agonists

The results are shown in fig. 1. 8-OHDPAT induced forepaw treading with an ED₅₀ of 8.0 μ mol/kg = 2.5 mg/kg s.c. Following simultaneous treatment with DOI (7 μ mol/kg = 2.5 mg/kg s.c.) the potency of 8-OHDPAT was increased approximately 20-fold. The ED₅₀ was 0.41 μ mol/kg = 0.13 mg/kg.

DOI did not induce typical forepaw treading when given alone but it did induce head twitches.

The latter were significantly inhibited by 8-OHDPAT in doses from 0.24 μ mol/kg, with an ED₅₀ value of 0.47 μ mol/kg = 0.15 mg/kg s.c.). Complete blockade was not obtained. Dose-response experiments with DOI showed that head twitches were induced with an ED₅₀ value of 0.51 μ mol/kg = 0.17 mg/kg s.c., while forepaw treading was absent. Forepaw treading was facilitated after combination of DOI with a threshold dose of 8-OHDPAT (1.9 μ mol/kg = 0.63 mg/kg s.c.) and an ED₅₀ of 0.27 μ mol/kg = 0.097 mg/kg was obtained with DOI. Head twitches were significantly inhibited by 8-OHDPAT at all dosages of DOI.

TFMPP was tested at two dose levels (1.0 and 4.2 μ mol/kg = 0.31 and 1.3 mg/kg, s.c.; data not shown). Neither head twitches nor forepaw treading were induced by TFMPP given alone. A slight increase in forepaw treading score followed the highest dose of TFMPP after combination with 8-OHDPAT (1.9 μ mol/kg). The scores after 8-OHDPAT and after 8-OHDPAT + TFMPP were 1.8 ± 0.63 and 3.3 ± 0.37 , respectively (mean $\pm$ S.E.M., n = 8, P = 0.05). The high dose of TFMPP inhibited the head twitch response to DOI (7.0 μ mol/kg, s.c.). These were 11 ± 1.2 and 6.3 ± 1.3 (mean $\pm$ S.E.M., n = 4, P = 0.03) head twitches after DOI and DOI + TFMPP, respectively.

3.3. Effects of antagonists

The effects of various antagonists on the response to combined treatment with 8-OHDPAT (1.9 μ mol/kg) and DOI (7.0 μ mol/kg) are shown

TABLE 1

Affinity of 5-HT agonists and antagonists for 5-HT_{1A}, 5-HT_{1B} and 5-HT₂ receptors. See methods for details.

	IC ₅₀ (nM)		
	[³ H]OHDPAT	[³ H]5-HT	[³ H]Ketanserin
<i>Agonists</i>			
8-OHDPAT	3.5	410	9200
DOI	11000	3000	110
TFMPP	460	97	360
<i>Antagonists</i>			
(–)Alprenolol	87	14000	6900
Ketanserin	5700	1500	2.0
Ritanserin	3800	910	0.40

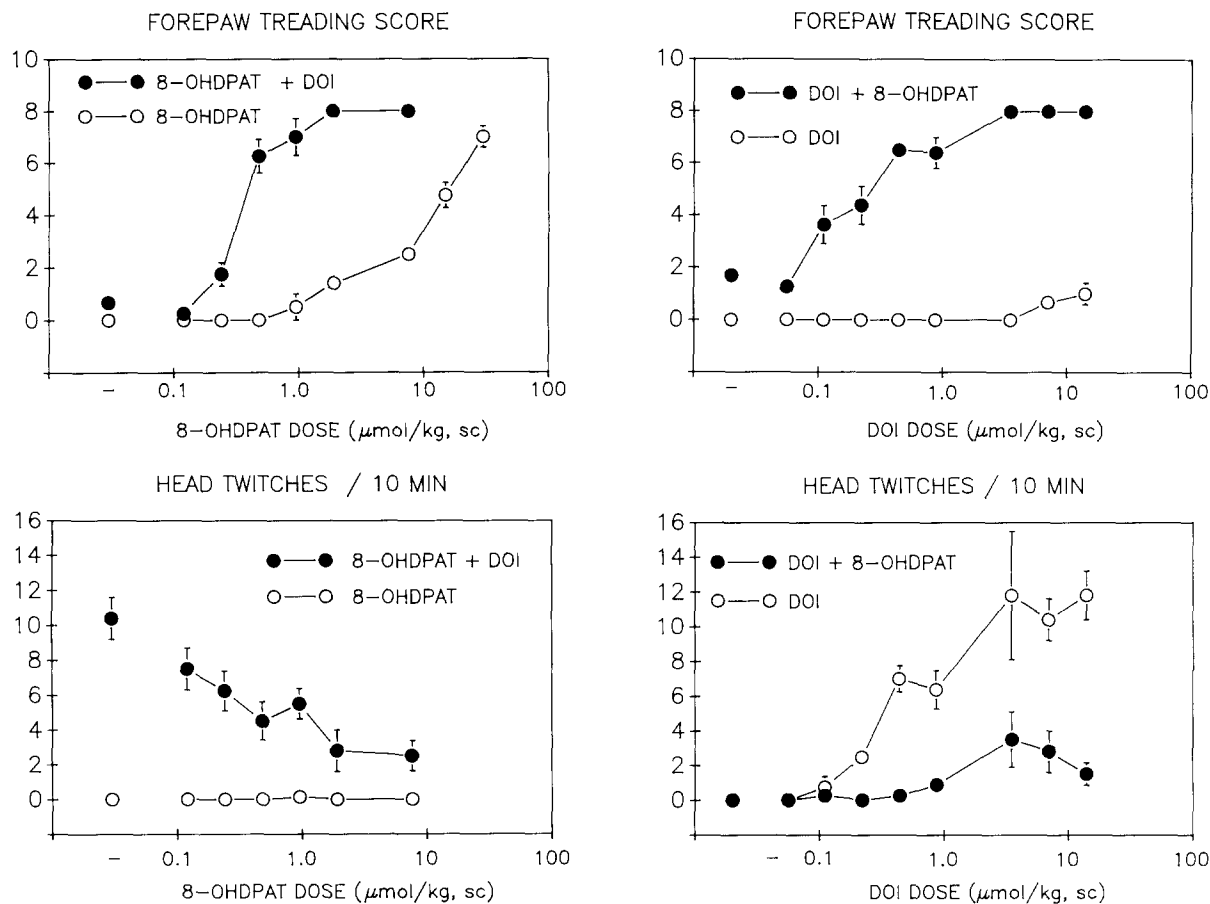


Fig. 1. Effects of 8-OHDPAT and DOI, alone and in combination, on forepaw treading and head twitches in rats. Left: 8-OHDPAT injected s.c. alone (○) or simultaneously with DOI (7 μmol/kg s.c.; ●). Forepaw treading (upper panel) was scored 15, 30, 45 and 60 min after injection as 0, 1 or 2 and the scores were added. The maximum score was thus 8. The effect of saline (○) and DOI (●) given alone is shown as single points to the left. Head twitches (lower panel) were counted in the same groups of rats from 30-40 min after injection. Right: DOI was injected alone (○) or simultaneously with 8-OHDPAT (1.9 μmol/kg s.c.; ●) and behaviour was evaluated as described above. Each point represents the mean ± S.E.M. of data from 4-8 rats. The control values are pooled results from a total of 12-16 rats.

in table 2. The 5-HT₂ antagonists, ketanserin and ritanserin, induced a dose-dependent inhibition of the forepaw treading response. The head twitch response to DOI + 8-OHDPAT was already low (see section 3.2) and was further decreased by ketanserin and ritanserin. Forepaw treading was also inhibited by (–)-alprenolol whereas the head twitches were slightly facilitated. The β-adrenoceptor antagonist, ICI 118.551, and the 5-HT₃

antagonist, ICS 205-930, had no effect on either forepaw treading or head twitches in comparison with the controls.

Finally, the effect of combined treatment with a non-effective dosage of (–)-alprenolol and either ketanserin or ritanserin was studied. Figure 2 shows that (–)-alprenolol increased the inhibitory effect of both compounds on forepaw treading, leading to almost complete blockade of the re-

TABLE 2

Effect of antagonists on forepaw treading and on head twitches induced by combined treatment with 8-OHDPAT and DOI. The antagonists were injected s.c. 30 min or 2 h (ritanserin) before combined treatment with 8-OHDPAT (1.9 μ mol/kg s.c.) and DOI (7.0 μ mol/kg). Parallel control groups treated with DOI alone had a forepaw treading score of 3.1% and 11.4 ± 0.89 head twitches ($n = 24$). 8-OHDPAT given alone induced a forepaw treading score of $32 \pm 6.1\%$. No head twitches were observed.

Pretreatment	Dose (μ mol/ kg s.c.) ^a	Forepaw treading % max. score (mean $\pm$ S.E.M.)	Head twitches counts/10 min (mean $\pm$ S.E.M.)
Saline	– (24)	100	2.9 ± 0.48
Ketanserin	0.40 (8)	88 ± 7.0	1.3 ± 0.37
	1.6 (12)	66 ± 4.6^c	0.58 ± 0.33^c
	6.3 (8)	17 ± 6.7^c	0^c
Ritanserin	0.33 (8)	61 ± 6.0^c	0.25 ± 0.25^c
	1.3 (4)	31 ± 3.6^c	0^b
(–)-Alprenolol	6.3 (8)	100	4.6 ± 1.3
	25 (12)	93 ± 6.2	5.7 ± 0.99
	50 (8)	38 ± 12^c	7.5 ± 1.2^d
ICI 118.551	64 (4)	100	0.25 ± 0.25
ICS 205-930	4.4 (4)	98 ± 2.0	1.8 ± 1.4

^a Number of animals shown in parentheses. ^b $P < 0.05$. ^c $P < 0.01$ significant inhibition compared with parallel control groups. ^d $P < 0.05$ significant increase compared with parallel control groups.

sponse. The head twitches were inhibited in all groups receiving ketanserin or ritanserin (data not shown).

4. Discussion

The results indicate that forepaw treading behaviour is facilitated by an interaction between 5-HT_{1A} and 5-HT₂ receptors.

Forepaw treading is part of the 5-HT syndrome and is the symptom which we find most reliable to score. Previous evidence suggests that 5-HT_{1A} receptors mediate this response (Tricklebank, 1985; Tricklebank et al., 1985a,b). This suggestion is consistent with our results showing that the maximum response is induced by the specific 5-HT_{1A} agonist 8-OHDPAT, whereas the preferential 5-HT_{1B} agonist, TFMPP, and the 5-HT₂ agonist,

DOI, are ineffective. However, co-treatment with 8-OHDPAT and DOI increased the potency of 8-OHDPAT markedly. This suggests that stimulation of 5-HT₂ receptors has a permissive role for the expression of the forepaw treading syndrome.

This may also explain why 5-HT₂ antagonists (e.g. ketanserin and ritanserin; Leysen and Gommeren, 1986) have some inhibitory effect on the forepaw treading syndrome induced by 8-OHDPAT or 5-methoxy-N,N-dimethyltryptamine, a non-selective 5-HT₁/5-HT₂ agonist (Tricklebank et al., 1985a,b; see also Goodwin and Green, 1985). Ketanserin and ritanserin also inhibited the response induced by DOI plus 8-OHDPAT (table 2). A similar inhibitory effect was found with the 5-HT₁ (and β -adrenoceptor) antagonist (–)-alprenolol (Middlemiss, 1986; table 1). Supplementary evidence for a 5-HT_{1A}/5-HT₂ interaction is provided by the marked synergistic effect of (–)-alprenolol and either ketanserin or ritanserin as inhibitors of forepaw treading (fig. 2). Involvement of β -adrenoceptors is unlikely since the β -adrenoceptor antagonist, ICI 118.551, was ineffective as in other models of 5-HT₁ receptor function

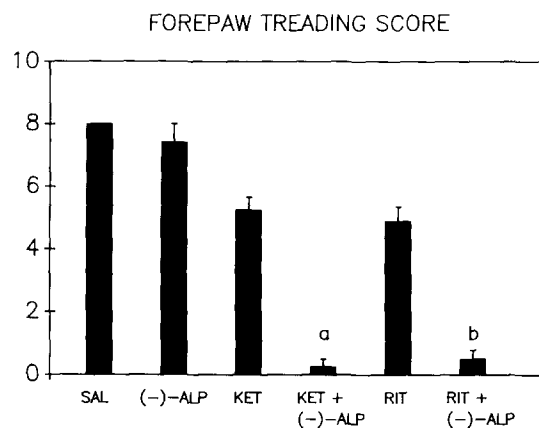


Fig. 2. Effects of antagonists alone or in combination on the forepaw treading induced by 8-OHDPAT plus DOI. Ketanserin (1.6 μ mol/kg s.c.) and (–)-alprenolol (25 μ mol/kg s.c.) were injected 30 min before, and ritanserin (0.33 μ mol/kg s.c.) was injected 2 h before 8-OHDPAT (1.9 μ mol/kg s.c.) plus DOI (7.0 μ mol/kg s.c.). Scoring was according to the legend to fig. 1. The results are the means $\pm$ S.E.M. from 4 (combination treatments) or 8 rats (other groups). ^a $P < 0.01$ significant inhibition compared with ketanserin and ^b $P < 0.01$ significant inhibition compared with ritanserin.

(Tricklebank et al., 1985b; Middlemiss, 1986). The 5-HT₃ receptor antagonist ICS 205-930 (Richardson et al., 1985; Kilpatrick et al., 1987) is also ineffective.

The interpretation of our results was complicated by recent observations showing that DOI has similar affinities for 5-HT₂ and 5-HT_{1C} sites (Hoyer and Karpf, 1988). Titeler et al. (1988), however, found a 40-fold higher affinity for 5-HT₂ than for 5-HT_{1C}. Moreover, ritanserin, which is considered to be a selective 5-HT₂ antagonist (Leysen and Gommeren, 1986), has a high 5-HT_{1C} affinity (Hoyer and Karpf, 1988). However, ketanserin shows at least 50-fold selectivity for 5-HT₂ versus 5-HT_{1C} receptors (Hoyer et al., 1985; Hoyer and Karpf, 1988). The potent inhibitory effect of ketanserin against the forepaw treading induced by DOI plus 8-OHDPAT thus favors 5-HT₂ rather than 5-HT_{1C} sites as the important mediator of the effect of DOI. This conclusion is further supported by the lack of effect of TFMPP which has a high 5-HT_{1C} affinity (Hoyer, 1988) and is active in an *in vivo* model suggested for 5-HT_{1C} activity (Kennett and Curzon, 1988).

The facilitatory effect of 5-HT₂ stimulation raises the question of whether the effect of 8-OHDPAT given alone depends on endogenous 5-HT activation of 5-HT₂ receptors. This would be analogous to similar findings from study of dopamine (DA) D-1 and D-2 receptors. It is known that endogenous activity at D-1 receptors or treatment with a D-1 agonist is necessary for the expression of the behavioural stimulation induced by a D-2 agonist. Following subchronic DA depletion D-1 and D-2 receptors become supersensitive and function independently of each other (for review, see Clark and White, 1987). The observation that the forepaw treading induced by 8-OHDPAT was preserved in rats treated for 3 days with an inhibitor of 5-HT synthesis (Tricklebank et al., 1985a) argues against the above hypothesis. However, adaptational increases in the sensitivity of 5-HT₁ receptors have been observed as early as 1 day after lesions of 5-HT neurones (Ortmann et al., 1981). This indicates that adaptational changes may also take place after the subchronic inhibition of 5-HT synthesis and thus influence the outcome of the behavioural experiment. Clearly further

work is needed before acceptance or rejection of these hypotheses.

The receptor interaction for the head twitch syndrome is different, since stimulation of 5-HT_{1A} receptors leads to inhibition of the response to a 5-HT₂ agonist. It cannot be excluded that the inhibition is caused by behavioural competition with forepaw treading, since the dose-response curves for the two responses mirror each other (fig. 1). A similar observation is that the inhibition of DOI-induced head twitches by 8-OHDPAT was partially reversed, in parallel to the inhibition of forepaw treading after treatment with (–)-alprenolol. However, preliminary results indicate that partial 5-HT_{1A} agonists such as buspirone and ipsapirone inhibit head twitches without inducing forepaw treading (Arnt, unpublished observations). An opposite 5-HT_{1A}/5-HT₂ interaction is also observed in the regulation of body temperature in rats (Gudelsky et al., 1986) and thus a similar balance model is possible for the head twitch syndrome.

The involvement of 5-HT_{1B} receptors in the expression of these behaviours is unlikely to be of importance. The preferential 5-HT_{1B} agonist TFMPP had a maximum effect at 5-HT_{1B} receptors at the doses used in the present study (Arnt, 1989) but had weak effects which showed similarities to those of 8-OHDPAT and DOI. However, since the selectivity of TFMPP for 5-HT_{1B} versus 5-HT_{1A} and 5-HT₂ receptors is limited (Glennon, 1987; present results, table 1), the weak effect of TFMPP may also depend on non-specific stimulation of 5-HT receptors.

Acknowledgements

We wish to thank Annette Aasberg, Jytte Pedersen, Sus Hansen, Brian Hansen and Tom Halborg for excellent assistance.

References

- Arnt, J., 1989, Characterization of the discriminative stimulus properties induced by 5-HT₁ and 5-HT₂ agonists in rats, *Pharmacol. Toxicol.* 64, 165.

- Clark, D. and F.J. White, 1987, Review: D1 dopamine receptor – The search for a function: A critical evaluation of the D1/D2 dopamine receptor classification and its functional implications, *Synapse* 1, 347.
- Glennon, R.A., 1987, Central serotonin receptors as targets for drug research, *J. Med. Chem.* 30, 1.
- Goodwin, G.M. and A.R. Green, 1985, A behavioural and biochemical study in mice and rats of putative selective agonist and antagonists for 5-HT₁ and 5-HT₂ receptors, *Br. J. Pharmacol.* 84, 743.
- Green, A.R. and D.G. Grahame-Smith, 1976, Effects of drugs on the processes regulating the functional activity of brain 5-hydroxytryptamine, *Nature* 260, 487.
- Gudelsky, G.A., J.I. Koenig and H.Y. Meltzer, 1986, Thermoregulatory responses to serotonin (5-HT) receptor stimulation in the rat. Evidence for opposing roles of 5-HT₂ and 5-HT_{1A} receptors, *Neuropharmacology* 25, 1307.
- Hjorth, S., A. Carlsson, P. Lindberg, D. Sanchez, H. Wikström, L.-E. Arvidsson, U. Hacksell and J.L.G. Nilsson, 1982, 8-Hydroxy-2-(di-n-propylamino)tetralin, 8-OH-DPAT, a potent and selective simplified ergot congener with central 5-HT-receptor stimulating activity, *J. Neural Transm.* 55, 169.
- Hoyer, D., 1988, Molecular pharmacology and biology of 5-HT_{1C} receptors, *Trends Pharmacol. Sci.* 9, 89.
- Hoyer, D., G. Engel and H.O. Kalkman, 1985, Molecular pharmacology of 5-HT₁ and 5-HT₂ recognition sites in rat and pig brain membranes: Radioligand binding studies with [³H]5-HT, [³H]8-OH-DPAT, (–)[¹²⁵I]iodocyanopindolol, [³H]mesulergine and [³H]ketanserin, *European J. Pharmacol.* 118, 13.
- Hoyer, D. and A. Karpf, 1988, [¹²⁵I]SCH 23982, a 'selective' D-1 receptor antagonist, labels with high affinity 5-HT_{1C} sites in pig choroid plexus, *European J. Pharmacol.* 150, 181.
- Kennett, G.A. and G. Curzon, 1988, Evidence that hypophagia induced by mCPP and TFMPP requires 5-HT_{1C} and 5-HT_{1B} receptors; hypophagia induced by RU 24969 only requires 5-HT_{1B} receptors, *Psychopharmacology* 96, 93.
- Kilpatrick, G.J., B.J. Jones and M.B. Tyers, 1987, Identification and distribution of 5-HT₃ receptors in rat brain using radioligand binding, *Nature* 330, 24, 746.
- Leysen, J.E. and W. Gommeren, 1986, Drug-receptor dissociation time, new tool for drug research: Receptor binding affinity and drug-receptor dissociation profiles of serotonin-S₂, dopamine-D₂, histamine-H₁ antagonists, and opiates, *Drug Dev. Res.* 8, 119.
- Lucki, I., S.M. Nobler and A. Frazer, 1984, Differential actions of serotonin antagonists on two behavioural models of serotonin receptor activation in the rat, *J. Pharmacol. Exp. Ther.* 228, 133.
- Middlemiss, D.N., 1986, Blockade of the central 5-HT autoreceptor by β -adrenoceptor antagonists, *European J. Pharmacol.* 120, 51.
- Niemegeers, C.J.A., F.C. Colpaert, J.E. Leysen, F. Awouters and P.A.J. Janssen, 1983, Mescaline-induced head-twitches in the rat: An vivo method to evaluate serotonin S₂ antagonists, *Drug Dev. Res.* 3, 123.
- Ortmann, R., S. Martin and P.C. Waldmeier, 1981, Supersensitivity to L-5-hydroxytryptophan after 5,7-dihydroxytryptamine injections in desmethylimipramine- and nomifensine-pretreated rats: Behavioral evidence for postsynaptic supersensitivity, *Psychopharmacology* 74, 109.
- Richardson, B.P., G. Engel, P. Donatsch and P.A. Stadler, 1985, Identification of serotonin M-receptor subtypes and their specific blockade by a new class of drugs, *Nature* 316, 126.
- Sloviter, R.S., E.G. Drust and J.D. Connor, 1978, Specificity of a rat behavioral model for serotonin receptor activation, *J. Pharmacol. Exp. Ther.* 206, 339.
- Titeler, M., R.A. Lyon and R.A. Glennon, 1988, Radioligand binding evidence implicates the brain 5-HT₂ receptor as a site of action for LSD and phenylisopropylamine hallucinogens, *Psychopharmacology* 94, 213.
- Tricklebank, M.D., 1985, The behavioural response to 5-HT receptor agonists and subtypes of the central 5-HT receptor, *Trends Pharmacol. Sci.* 6, 403.
- Tricklebank, M.D., C. Forler and J.R. Fozard, 1985a, The involvement of subtypes of the 5-HT₁ receptor and of catecholaminergic systems in the behavioural response to 8-hydroxy-2-(di-n-propylamino)tetralin in the rat, *European J. Pharmacol.* 106, 271.
- Tricklebank, M.D., C. Forler, D.N. Middlemiss and J.R. Fozard, 1985b, Subtypes of the 5-HT receptor mediating the behavioural responses to 5-methoxy-N,N-dimethyltryptamine in the rat, *European J. Pharmacol.* 117, 15.